Chapter - 6

STUDIES ON ANTIOXIDANT ACTIVITY OF THE LEAVES OF *BISCHOFIA JAVANICA* AND *FRAXINUS FLORIBUNDA*

6.1 Introduction:

Oxygen free radical induced cellular damage has been implicated in many pathological conditions, like malignancy, ageing process, inflammation and degenerating diseases. ^[1-2] Reactive oxygen species (ROS) have been known to cause tissue injury through covalent bonding and lipid peroxidation.^[3] Antiinflammatory drugs could conceivably affect oxidant damage at the sites of inflammation in several ways. They might directly scavenge such reactive oxidants as hydroxyl radical (OH) and hypochlorous acid (OHCl). ^[4] It has been shown in most cases, if not all, antiinflammatory drugs are capable of reacting quickly with OH radical. Hence, any antiinflammatory drug present at a site of inflammation would remove OH radical. Antioxidants act as major defense against radical-mediated toxicity by protecting the damages caused by free radicals. Plant-derived antioxidants may function as reducing agents, scavengers of free radicals and metal ion chelators, among others. ^[5] The antioxidant activity in plasma has been shown to increase after consumption of foods high in antioxidants. ^[6] Thus, phytochemicals may combat oxidative stress in human body by maintaining a balance between oxidants and antioxidants

6.1.1 DPPH Radical Scavenging Assay:

The 1, 1-diphenyl- 2picryl hydrazyl (DPPH) radical was widely used a reagent to evaluate free radical scavenging activity of antioxidants. DPPH is a stable free radical and accepts an electron or hydrogen atom to become a stable diamagnetic molecule. ^[7-8] The reduction capability of DPPH radical is determined by the decrease in absorbance induced by antioxidants.

6.1.2 Lipid Peroxidation Assay:

Lipid peroxidation is the oxidative degradation of polyunsaturated fatty acids and involves formation of lipid radicals leading to membrane damage. Free radicals induce lipid peroxidation in poly unsaturated lipid rich areas like brain and liver. Malondialdehyde (MDA), the secondary product of lipid peroxidation, is estimated in the plasma samples utilizing the colorimetric reaction with thiobarbituric acid (TBA). ^[9] It gives the index of the extent of lipid peroxidation. Since the assay estimates the amount of TBA reactive substance e.g. MDA, it is also known as TBARS (thiobarbituric acid reactive substance) test.

6.1.3 Hydroxyl Radical Scavenging Assay:

The hydroxyl radical scavenging activity is measured as the percentage of inhibition of hydroxyl radicals generated in the Fenton's reaction mixture, by studying the competition between deoxy ribose and the test drug for hydrogen radicals generated from $Fe^{3+}/ascorbate/EDTA/H_2O_2$ systems. The hydroxyl radicals attack deoxy ribose which eventually results in TBARS formation.^[10]

6.2 Instruments, Chemicals and Drugs:

UV-Visible double beam spectrophotometer (Shimadzu Corporation, Japan), cold centrifuge are available in the laboratory of Himalayan Pharmacy Institute. DPPH (1,1-diphenyl-2-picryl hydrazyl), Tris-Hcl, Ferrous sulphate, Ascorbic acid, Trichloroacetic acid (TCA), Thiobarbituric acid (TBA), Potassium dihydrogen phosphate-pottasium hydroxide buffer, Deoxy ribose, α -tocopherol, EDTA, Ferric chloride and hydrogen peroxide were purchased from Sigma and other chemicals were procured from local firms and were of analytical grade.

6.3 Animals:

Wistar female albino rats (150-250g) were used in this study. They were housed in large propylene cages and kept at $22\pm2^{\circ}$ C in 12 h dark-light cycle. The animals were fed with pellet food and water *ad libitum*. All animals were acclimatized to the laboratory environment for at least one week before the experimental session. The animal experiment was approved by Institutional Animal Ethical Committee (IAEC) *No: HPI/07/60/IAEC/0006*.

6.4 Antioxidant Studies of the leaves of *Bischofia javanica*6.4.1 DPPH Radical Scavenging Assay:

To different concentrations (20, 40, 80,160 and 320µg/ml) of methanol extract of the leaves of *B.javanica* and its isolated compound, Friedeline-3α-acetate in methanol, 1ml of a methanolic solution of 0.2Mm DPPH ^[11] was added. After mixing thoroughly, the mixture was allowed to stand in dark for 30min and the absorbance at 523 nm was measured using methanol for the baseline correction. The results were then compared with that of the control prepared in the same manner but without the sample. Radical scavenging activity was calculated and expressed in percentage, α-tocopherol at the concentration range of 20, 40, 80, 160, 320 µg/ml was used as a positive control for each concentration. Readings were taken in quadruplicate. The antioxidant activity was expressed as IC₅₀ value, which is defined as the concentration (µg/ml) of extract required to inhibit the formation of DPPH radicals by 50%.

6.4.2 Lipid Peroxidation Assay:

The brains of normal female albino rats of the Wistar strain (age 3months) were dissected and homogenized in chilled 20mM Tris-HCl, pH 7.4. The homogenate was centrifuged at 20,000×g for 15 min at 4°C. A 0.5-ml aliquot of the supernatant was then taken to incubate with 0.3 ml of the test sample of methanol extract of leaves of *B.javanica* (BJ) and Friedelin-3α-acetate at the concentrations of 20, 40, 80,160 and 320µg/ml), 0.1 ml of 10 mM FeSO4, and 0.1 ml of 0.1mM ascorbic acid at 37°C for 1h. The reaction was then stopped by addition of 0.75 ml of 10% solution of trichloroacetic acid (TCA) and 0.5 ml of 1% (w/v) solution of thiobarbituric acid (TBA). The mixture was then heated at 100°C for 45 min. After centrifugation, the precipitated proteins were removed and the color of the malondialdehyde (MDA)-TBA complex in the supernatant was measured at 532 nm ^[12] in Uv-Vis spectrophotometer. α-tocopherol at the concentrations of 20,40,80,160 and 320 µg/ml was used as a positive control. For each concentration readings were taken in quadruplicate.

6.4.3 Hydroxyl Radical Scavenging Assay:

The hydroxyl radical scavenging activity of methanol extract of *B.javanica*, its pure compound, Friedelin- 3α -acetate and the reference compound α -tocopherol was determined by the method of Aruoma et al.^[13] The assay mixture, in a total volume of 1.2 ml, contained deoxyribose (2.8mM), FeCl₃ (25mM), EDTA (100mM), H₂O₂ (2.8mM), KH₂PO₄/KOH buffer, pH 7.4 (10mM), various concentrations of test drugs and standard (20-320µg/ml) and the pH was then readjusted to 7.4. The ascorbate (100mM) was then added to start the reaction. After incubation at 37 °C for 1 h, 1 ml of 1% (w/v) thiobarbituric acid (TBA) in 50mM NaOH and 1ml of 2.8% (w/v) trichloroacetic acid (TCA) was added to the reaction mixture and placed in a hot water bath maintained at 80°C for up to 20 min. The contents were cooled, centrifuged and absorbance of the supernatant was read at 532 nm against the control. Readings were taken in quadruplicate.

6.5 Antioxidant Studies of the Leaves of *Fraxinus floribunda*6.5.1 DPPH Radical Scavenging Assay:

To different concentrations (20, 40, 80,160 and 320µg/ml) of methanol extract of the leaves of *F.floribunda* and its isolated compounds β -amyrin in methanol, 1ml of a methanolic solution of 0.2Mm DPPH ^[11] was added. After mixing thoroughly, the mixture was allowed to stand in dark for 30min and the absorbance at 523 nm was measured using methanol for the baseline correction. The results were then compared with that of the control prepared in the same manner but without the sample. Radical scavenging activity was calculated and expressed in percentage, α -tocopherol at the concentration range of 20, 40, 80, 160, 320 µg/ml was used as a positive control for each concentration. Readings were taken in quadruplicate. The antioxidant activity was expressed as IC₅₀ value, which is defined as the concentration (µg/ml) of extract required to inhibit the formation of DPPH radicals by 50%.

6.5.2 Lipid Peroxidation Assay:

The brains of normal female albino rats of the Wistar strain (age 3months) were dissected and homogenized in chilled 20mM Tris-HCl, pH 7.4. The homogenate was centrifuged at 20,000×g for 15 min at 4°C. A 0.5-ml aliquot of the supernatant was then taken to incubate with 0.3 ml of the test sample of methanol extract of leaves of *Fraxinus floribunda* and β-amyrin at the concentrations of 20, 40, 80,160 and 320µg/ml), 0.1 ml of 10 mM FeSO4, and 0.1 ml of 0.1mM ascorbic acid at 37°C for 1h. The reaction was then stopped by addition of 0.75 ml of 10% solution of trichloroacetic acid (TCA) and 0.5 ml of 1% (w/v) solution of thiobarbituric acid (TBA). The mixture was then heated at 100°C for 45 min. After centrifugation, the precipitated proteins were removed and the color of the malondialdehyde (MDA)-TBA complex in the supernatant was measured at 532 nm ^[12] in UV-Vis spectrophotometer. α -tocopherol at the concentrations of 20,40,80,160 and 320 µg/ml was used as a positive control. For each concentration readings were taken in quadruplicate.

6.5.3 Hydroxyl Radical Scavenging Assay:

The hydroxyl radical scavenging activity of methanol extract of *F.floribunda*, its isolated pure compound, β -amyrin and the reference compound α -tocopherol was determined by the method of Aruoma et al. ^[13] The assay mixture, in a total volume of 1.2 ml, contained deoxyribose (2.8mM), FeCl₃ (25mM), EDTA (100mM), H₂O₂ (2.8mM), KH₂PO₄/KOH buffer pH 7.4 (10mM), various concentrations of test drugs and standard (20–320µg/ml) and the pH was then readjusted to 7.4. The ascorbate (100mM) was then added to start the reaction. After incubation at 37 °C for 1 h, 1 ml of 1% (w/v) thiobarbituric acid (TBA) in 50mM NaOH and 1ml of 2.8% (w/v) trichloroacetic acid (TCA) was added to the reaction mixture and placed in a hot water bath maintained at 80°C for up to 20 min. The contents were cooled, centrifuged and absorbance of the supernatant was read at 532 nm against the control. Readings were taken in quadruplicate.

6.6 Results of the Antioxidant Studies of the Leaves of *Bischofia javanica*:6.6.1 DPPH Radical Scavenging Assay:

The percentage of DPPH radical scavenging activity was calculated by using the formula ----

Absorbance of Control - Absorbance of sample

Scavenging (%) =

Absorbance of Control

×100

The test drugs, methanol extract of *Bischofia javanica* (BJ) and Friedelin-3 α -acetate shown to have concentration dependent antioxidant activities. The IC₅₀ value of BJ was found to be 118.29 µg/ml, for the isolated compound, Friedelin-3 α -acetate it was188.08 µg/ml (Table 6.1 and Figure 6.1, 6.2, 6.3). The scavenging activity of BJ and Friedelin-3 α -acetate at 320µg/ml was found to be 84.26 and 68.21% respectively. The slope of the curve showed the precision of the linearity of concentration dependent radical scavenging activity in the Figures 6.1 to 6.3.

6.6.2 Lipid Peroxidation Assay:

The percentage of scavenging activity of the drugs was calculated using the formula given in 6.6.1. The results of the study revealed that the antioxidant activities of the test drugs are more in the concentration range of 160-320 µg/ml. The coloration produced by the Thio Barbituric Acid Reactive Substance (TBARS) was reduced by the test drug in between these concentrations in remarkable manner. The IC₅₀ values of the drugs BJ, Friedelin-3a-acetate and a-tocopherol were 142,191.10 and 135.31 µg/ml (**Tables 6.2**) respectively. Linear regression curve depicted the R² of Friedelin-3a-acetate is 0.9713 which is much more than that of BJ (0.9184) and a-tocopherol (0.9126) as shown in the **Figures 6.4, 6.5 and 6.6**.

6.6.3 Hydroxyl Radical Scavenging Assay:

The percentage of hydroxyl radical scavenging activity was determined using the formula given in **6.6.1**. The IC₅₀ value of hydroxyl radical scavenging activity of the test drugs are found to be higher compared to DPPH and Lipid peroxidation assay (Table 6.3). The free radical (OH) scavenging activity for the test drugs are found to be concentration dependent as shown in the **Figure 6.7, 6.8 and 6.9**. In the concentration range of 20-320µg/ml of test drugs, the IC₅₀ for the test drugs BJ and friedelin-3α-acetate was found to be 185.23 and 163.37µg/ml respectively and for the standard drug α-tocopherol it was 158.95µg/ml.

6.7 Results of Antioxidant Studies of the Leaves of Fraxinus floribunda:

6.7.1 DPPH Radical Scavenging Assay:

The percentage of DDPH radical scavenging activity was calculated by using the formula---

In this study the DPPH radical scavenging activity test drugs was found to be concentration dependent in the concentration range of $20-320\mu g/ml$. β -amyrin, the pure compound isolated from the leaves of *Fraxinus floribunda* (FF) shown to have antioxidant activity (72.14%) better than the methanol extract (61.42%) of the leaves (**Table 6.4**). The IC₅₀ values of FF, β -amyrin and α -tocopherol were determined by Linear regression analysis as shown in the **Figures 6.10, 6.11 and 6.12**.

6.7.2 Lipid Peroxidation Assay:

The percentage of scavenging activity of the drugs was calculated using the formula given in 6.7.1. The results of the Lipid peroxidation assay revealed that the antioxidant activity of the test drugs increase on increasing the concentration of the drugs. The isolated phytoconstituent β -amyrin showed better antioxidant activity (IC₅₀137.92µg/ml)

compared to FF (IC₅₀ 172.52 µg/ml and α -tocopherol (IC₅₀ 149.13µg/ml) (**Table 6.5**). The linear regression curve of FF, β -amyrin and α -tocopherol are represented in the Figures 6.13, 6.14 and 6.15.

6.7.3 Hydroxyl Radical Scavenging Assay:

The percentage of hydroxyl radical scavenging activity was determined using the formula given in 6.7.1. The results of the hydroxyl radical scavenging activity assay are presented in the **Table 6.6** and **Figures 6.16, 6.17 and 6.18**. The IC₅₀ value of methanol extract of *F.Floribunda* was found to be more (166.63µg/ml) than that of standard drug α -tocopherol (149.15µg/ml) while the isolated compound, β -amyrin showed better activity (129.75µg/ml) than both FF and α -tocopherol.

Table 6.1 Antioxidant activity of leaves of Bischofia javanica by DPPHradical scavenging assay

Drug	Concentration (µg/ml)	Scavenging activity (%)	IC ₅₀ (μg/ml)
BJ	20	23.42±1.64	118.29
	40	32.56±1.56	
	80	46.15±2.84	
	160	69.23±3.32	
	320	84.26±2.11	
Friedeline-	20	18.45±3.41	188.08
3α-acetate	40	29.24±2.98	
	80	36.86±3.42	
	160	48.24±2.59	
	320	68.21±4.36	
a-tocopherol	20	20.22±3.12	146.28
	40	37.51±3.23	
	80	41.19±1.14	
	160	53.20±4.56	
	320	78.67±1.64	

Each value represents means of quadruplicate determinations \pm SD,

BJ - Methanol extract of Bischofia javanica

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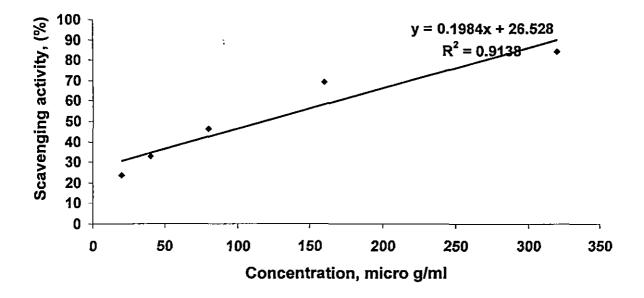


Figure 6.1 Linear regression analysis of antioxidant activity of methanol extract of *Bischofia javanica* by DPPH assay

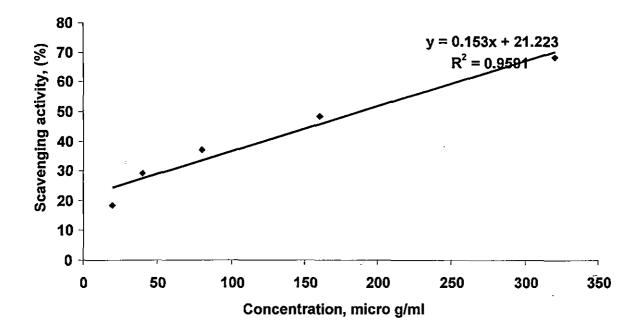


Figure 6.2 Linear regression analysis of antioxidant activity of Friedelin-3alpha-acetate by DPPH assay

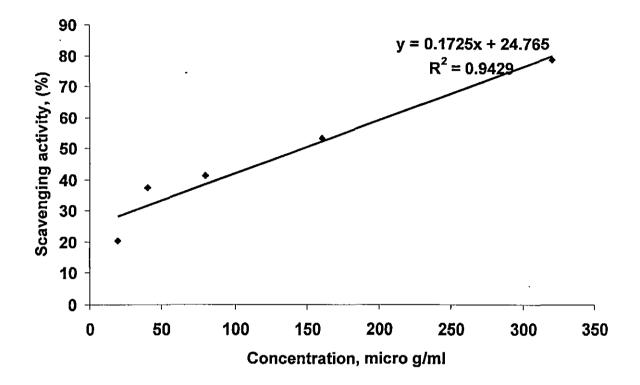


Figure 6.3 Linear Regression analysis of antioxidant activity of alphatocopherol by DPPH assay

Table 6.2 Antioxidant activity of leaves of Bischofia javanica by Lipid peroxidation assay

Drug	Concentration	Scavenging activity (%)	IC ₅₀ (μg/ml)
	(µg/ml)		
BJ	20	16.11±2.43	
	40	37.06±2.76	
	80	43.14±3.42	142.00
	160	55.23±1.32	· ·
	320	81.43±2.34	
Friedeline-3a-	20	12.10±1.45	
acetate	40	20.26±2.23	191.10
	80	32.57±1.65	
	160	48.19±3.78	
	320	72.13±1.96	
α-tocopherol	20	10.18±2.45	135.31
	40	27.43±1.43	
	80	42.67±2.13	
	160	67.20±4.21	
	320	88.66±3.79	

Each value represents means of quadruplicate determinations \pm SD, BJ - Methanol extract of *Bischofia javanica*

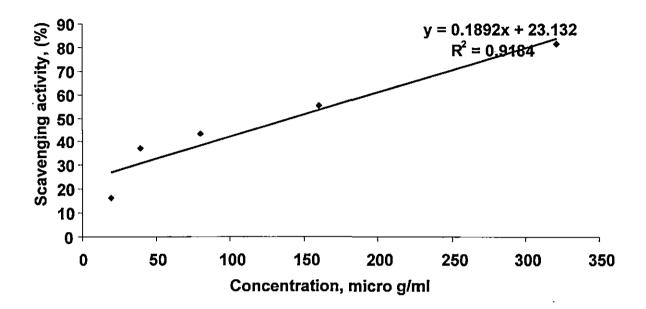


Figure 6.4 Linear regression analysis of antioxidant activity of methanol extract of leaves of *Bischofia javanica* by Lipid peroxidation assay

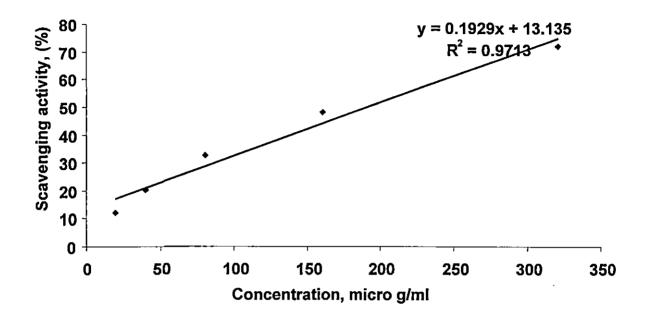


Figure 6.5 Linear regression analysis of antioxidant activity of Friedelin-3alpha-acetate by Lipid peroxidation assay

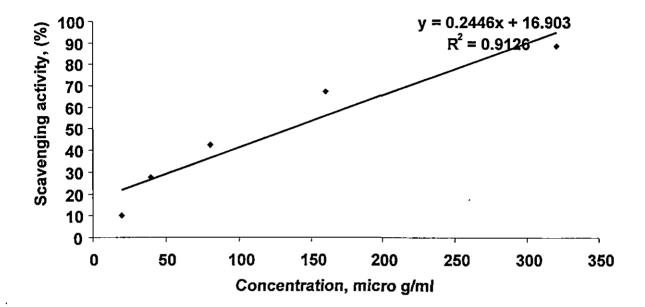


Figure 6.6 Linear regression analysis of antioxidant activity of alphatocopherol by Lipid peroxidation assay

Table 6.3 Antioxidant activity of leaves of Bischofia javanica by Hydroxyl radical scavenging assay

Drug	Concentration	Percentage of scavenging	IC ₅₀ (μg/ml)
	(µg/ml)		
BJ	20	10.01±2.21	185.23
	40	28.19±3.42	
	80	35.25±1.23	
	160	48.26±1.43	
	320	72.14±5.14	
Friedeline-3a-	20	12.17±4.67	163.37
acetate	40	23.32±1.23	
	80	38.67±3.45	
	160	55.19 ± 2.31	
	320	79.11±1.97	
a-tocopherol	20	17.18±2.87	158.95
	40	23.46±3.56	
	80	43.57±4.57	
	160	69.20±1.23	
	320	84.79±2.54	

Each value represents means of quadruplicate determinations \pm SD, BJ – Methanol extract of *Bischofia javanica*

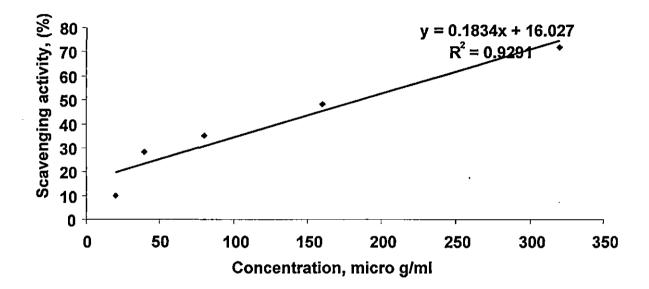


Figure 6.7 Linear regression analysis of antioxidant activity of methanol extract of leaves of *Bischofia javanica* by Hydroxy radical scavenging assay

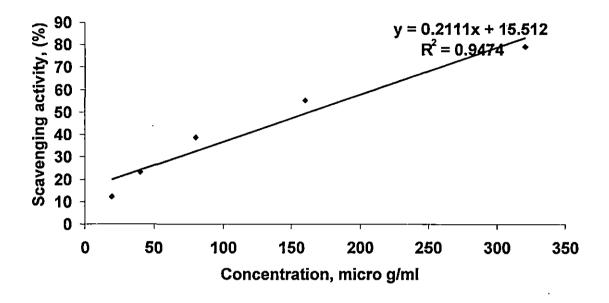


Figure 6.8 Linear regression analysis of antioxidant activity of Friedelin-3alpha-acetate by hydroxyl radical scavenging assay

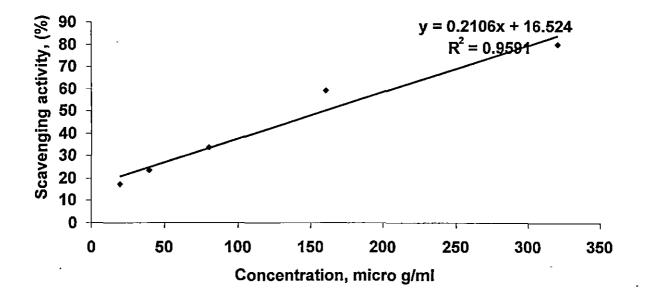


Figure 6.9 Linear regression analysis of antioxidant activity of alphatocopherol by hydroxyl radical scavenging assay

Drug	Concentration (µg/ml)	Scavenging activity(%)	IC ₅₀ (μg/ml)
FF	20	12.43±2.31	218.77
	40	26.98±3.21	
	80	38.32±1.56	
	160	44.15±2.98	
	320	61.42±1.44	
β-amyrin	20	14.54±0.98	168.47
	40	29.67±1.23	
	80	40.97±2.87	
	160	54.26±1.29	
	320	72.14±6.14	
a-tocopherol	20	18.25±2.78	154.68
	40	34.57±2.44	
	80	39.11±1.67	
	160	56.01±2.90	
	320	75.54±1.43	

Table 6.4 Antioxidant activity of leaves of Fraxinus floribunda by DPPHreduction assay

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Each value represents means of quadruplicate determinations \pm SD,

FF - Methanol extract of Fraxinus floribunda

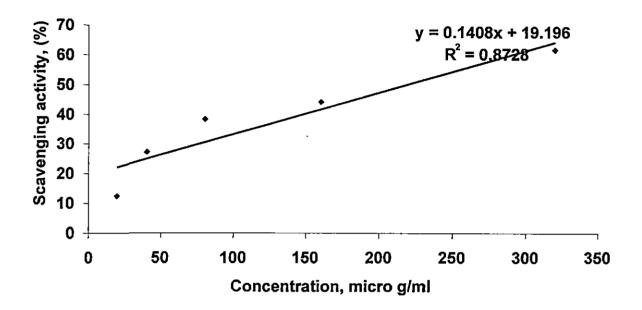


Figure 6.10 Linear regression analysis of antioxidant activity of methanol extract of leaves of *Fraxinus floribunda* by DPPH assay

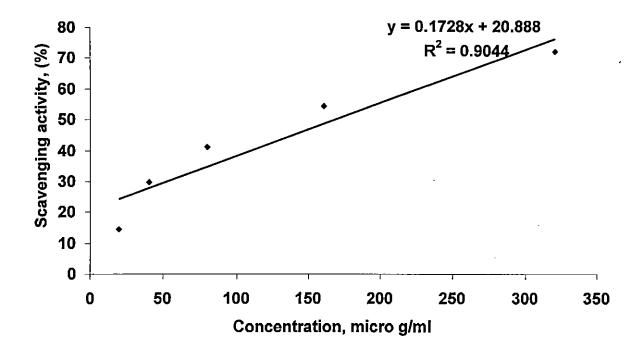
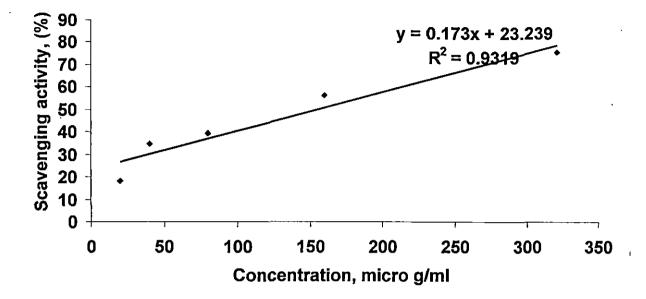


Figure 6.11 Linear regression analysis of antioxidant activity of Beta amyrin by DPPH assay



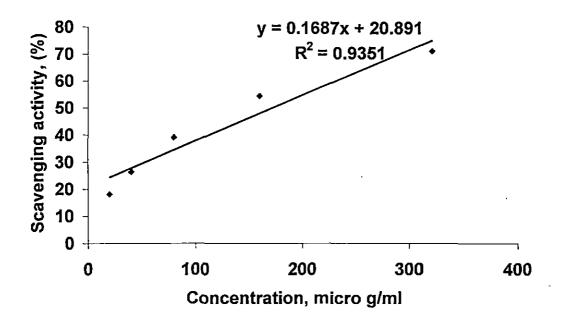
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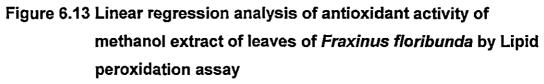
Figure 6.12 Linear regression analysis of antioxidant activity of alphatocopherol by DPPH assay

Drug	Concentration (µg/ml)	Scavenging activity (%)	IC ₅₀ (µg/ml)
FF	20	18.14±3.78	
	40	26.45 ±2.88	
	80	39.16±1.43	172.54
	160	54.27±2.14	
	320	71.00±2.72	
β-amyrin	20	14.45±1.98	137.92
	40	28.29±1.78	
	80	46.27±3.11	
	160	65.32±2.41	
	320	81.23±1.10	
a-tocopherol	20	12.32±1.08	149.13
	40	24.43±3.18	
	80	46.67±2.46	
	160	62.20±2.67	
	320	78.66±2.34	

Table 6.5 Antioxidant activity of leaves of Fraxinus floribunda by Lipidperoxidation assay

Each value represents means of quadruplicate determinations \pm SD, FF- Methanol extract of *Fraxinus floribunda*





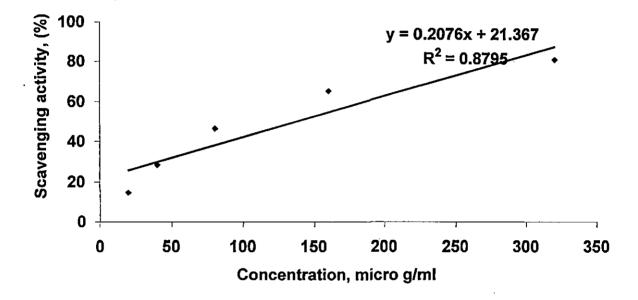
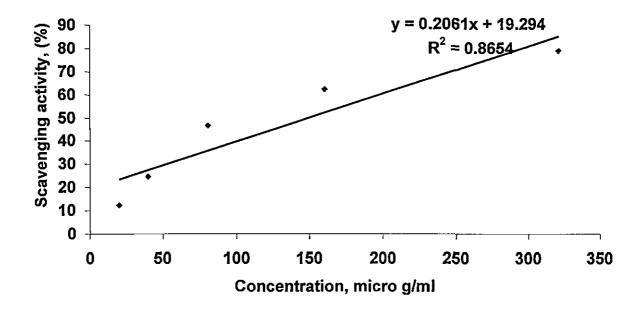


Figure 6.14 Linear regression analysis of antioxidant activity of Betaamyrin by Lipid peroxidation assay



Figure[•]6.15 Linear regression analysis of antioxidant activity of Alphatocopherol by Lipid peroxidation assay

Drug	Concentration (µg/ml)	Percentage of scavenging	IC ₅₀ (µg/ml)
FF	20	11.23±1.98	166.63
	40	23.87±2.64	
	80	41.76±3.21	
	160	58.91±1.76	
	320	73.21±1.33	
B-amyrin	20 .	19.24±1.27	129.85
	40	29.36±2.24	
	80	48.57±2.95	
	160	65.49±1.31	
	320	81.57±3.07	
a-tocopherol	20	15.16±2.87	149.13
	40	28.76±1.66	
	80	47.37±2.59	
	160	68.20±2.23	
	320	81.99±1.55	

Table 6.6 Antioxidant activity of leaves of Fraxinus floribunda by Hydroxylradical scavenging assay

Each value represents means of quadruplicate determinations \pm SD, FF - Methanol extract of *Fraxinus floribunda*

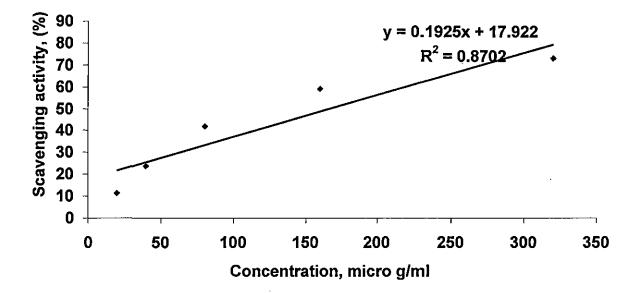


Figure 6.16 Linear regression analysis of antioxidant activity of methanol extract of leaves of *Fraxinus floribunda* by hydroxyl radical scavenging assay

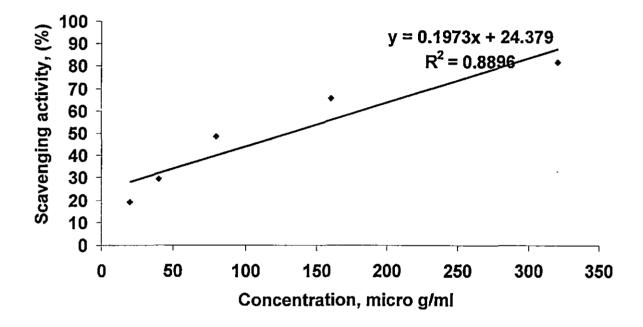


Figure 6.17 Linear regression analysis of antioxidant activity of Betaamyrin by hydroxyl radical scavenging assay

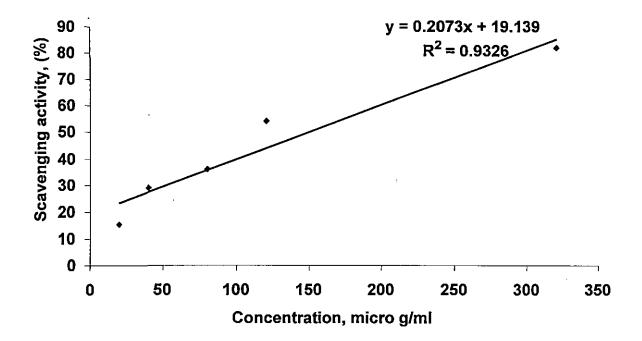


Figure 6.18 Linear regression analysis of antioxidant activity of Alphatocopherol by hydroxyl radical scavenging assay

References:

- 1. Ames BN, Shigenaga MK, Hagen TM. Oxidants, antioxidants and the degenerative diseases of ageing. Proc Natl Acad Sci 1993;90:7915.
- 2. Maxwell SRJ. Prospects for the use of antioxidant therapies. Drugs 1995;49:345.
- Geesin JG, Gordon JS, Berg RA. Retinoids affect collagen synthesis through inhibition of ascorbate-induced lipid peroxidation in cultured human dermal fibroblasts. Arch Biochem Biophys 1990;278:352.
- Aruoma OI, Halliwell B. The iron-binding and hydroxyl radical scavenging action of anti-inflammatory drugs. Xenobiotica 1994;18:459.
- 5. RiceEvans CA, Miller NJ, Paganga G. Structure antioxidant, activity relationships of flavonoids and phenolic acids. Free Rad Biol Med 1996;20:933-956.
- 6. Temple NJ. Antioxidants and disease: more questions than answers. Nutrition Research 2000;20:449-459.
- Oyaizu M. Studies on product of browning reaction prepared from glucose amine. Jap J Nutri 1986;44:307-315.
- 8. Soares JR, Dins TCP, Cunha AP, Ameida LM. Antioxidant activity of some extracts of *Thymus zygis*. Free Rad Res 1997;26:469-478.
- 9. Placer ZA, Crushman LL, Johnson BC. Estimation of products of lipid peroxidation (Malonyldialdehyde) in biochemical systems. J Biol Chem 1966;16:359.

- Hagerman AE, Riedl KM, Jones GA, Sovik KN, Ritchard NT, Hartzfeld PW. High molecular weight plant polyphenolics (Tannins) as biological antioxidants. J Agri Food Chem 1998;46:1887-1892.
- Yen GC, Hsieh GL .Antioxidant effects of dopamine and related compounds. Biosci Biotech Biochem 1997;61:1646-1649.
- 12. Lui F, Ng TB. Antioxidative and free radical scavenging activities of selected medicinal herbs. Life Sci 2000;66:725-735.
- Auroma OI. Deoxyribose assay for detecting hydroxyl radicals. Meth Enzymol 1994;233:57-66.